

Assessment of Polycyclic Aromatic Hydrocarbons (PAHs) in Commonly Consumed Shellfish from Kula, Rivers State, Nigeria

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Abstract

Polycyclic Aromatic Hydrocarbons (PAHs) were assessed in shellfishes (whelk, oyster and periwinkle) from Kula, Rivers State, Nigeria. The PAHs determination was done using gas chromatography (GC) coupled with flame ionization detector (FID) (Hewlett Packard, Wilmington, DE, USA), powered with HP chemstation Rev. A09:01 (10206) software. Human health risk assessment models based on United States Environmental Protection Agency (USEPA) was used to characterize risks of PAHs exposure to non cancer (Hazard Index) while and excess cancer risk (ECR). From the results, Benzo [a] Anthrancene (BaA) had highest concentrations in whelk (0.689±0.003) and Periwinkle (0.930±0.001) while Naphthalene had highest concentration in oyster (2.000±0.000). The Total concentration of PAHs in µ g/kg for whelk, oyster and periwinkle were 1.797±0.013, 3.977 ±0.024 and 1.564±0.017 while the estimated daily intake (EDI) of PAHs (mg/kg/day) via consumption of shell fish ranged from 2.00×10^{-4} to 6.40×10^{-2} , 7.0×10^{-4} to 1.86×10^{-1} and 0 to 8.64×10^{-2} far above oral reference dose (RFD) respectively. The toxic equivalents (TEQs) values were 1.276x10⁻⁴, 1.252x10⁻⁴ and 4.034x10⁻⁴ for whelk, oyster and periwinkle respectively, were significantly (p<0.05) higher than the screening value (SV) for shellfish 1.81×10^{-5} mg/kg. The estimated excess cancer risk (ECR) obtained for whelk was (3.0×10^{-4}) , oyster (2.00×10^{-4}) and periwinkle (3.24×10^{-4}) . These values were far above the USEPA acceptable (1×10^{-4}) . From this study, it can be deduced that bioaccumulation of PAHs in the shellfish is a potential health hazard to consumers. Carcinogenic indices indicated that daily Intake of contaminated



shellfishes exposures the local populace to cancer risks.

Keywords: Kula, PAHs, Shellfish, Health risk, Crude oil

1. Introduction

Sea foods are aquatic organisms that serve as major source of proteins to coastal communities around the globe (Serge and Andrew, 2018). They are further divided into fish and shell fish. Most of the shell fish or shell fishes are obtained from salt water environments and have been regarded as good bioaccumulation and bio-indicator (Onyema, 2018). Several of them have been used to monitor different classes of pollutants in the aquatic environment (Ueno *et al.*, 2003; Balogun *et al.*, 2011; Andem *et al.*, 2013).

Polycyclic aromatic hydrocarbons (PAHs) are persistent organic pollutants in the environment which are emitted from both natural and anthropogenic activities (Nkpaa et al., 2015). There are over 100 different chemical groups from PAHs emission that have ability to travel long distance resist biodegradation in the environment. They also have the ability to bio-accumulate in living organisms (Poster et al., 2006; Orish et al., 2015; Tongo et al., 2017). PAHs are listed as priority pollutants by the United States Environmental Protection Agency (USEPA, 2000) and the European Union (EU, 2014) because they are linked to environmental and health issues (Tongo et al., 2017). PAHs can find their way into the marine environment through petroleum pollution, fallout from air and effluents from industrial/sewage treatment plants settle at sediments of estuaries, bio-accumulate in sea organisms and passed to humans through the food chain with high degree of toxicity (Esra, 2016). In recent times a number of environmental agencies and scientific institutions have paid much attention to the presence of PAHs in the environmental (Tavakoly Sany et al., 2014; Zahra et al., 2014) and the potential to cause varying adverse effects on human health. Some PAHs are considered to be mutagenic and/or carcinogenic. The metabolism of PAHs requires the activation of numerous enzymes of the cytochrome p450 oxidase system involved in epoxidation and conjugation. This reaction that can lead to depletion in antioxidant enzymes and induction of oxidative stress leading to cataracts, kidney and liver damage, setting the stage for rapid aging and death of cells (Singh et al., 2008; Androutsopoulos et al., 2009).

Health risk assessment is defined as the process that evaluates the toxic properties of chemical substances and the effects upon human exposure (Nkpaa *et al.*, 2015). The risk assessment is a multi-step procedure that comprises data collection (gathering and analyzing the site data relevant to human health); exposure assessment (estimation of the potential human exposures); toxicity assessment (determination of adverse effects associated with exposure) and risk characterization (summarizes and combines outputs of the calculations of exposure and toxicity assessments) (EPA, 2004; Li *et al.*, 2010).

Health risk assessment (HRA) for PAHs in sea foods have been strongly encouraged by different environmental agencies (Zelinkova and Wenzi, 2015). So far, there is limited information on health risk assessment of PAHs in sea foods consumed by the local populace in Kula Kingdom. Therefore, this study is aimed at estimating the levels of some PAHs in



shellfish (*Busycon carica* (whelk,) *Crassostrea gigas* (oyster), *Tympanotonos fuscatus* (periwinkle)) in the study area and to use the Human Health Risk Assessment model to characterize and evaluate the potential health risk upon expose via ingestion route.

2. Materials and Methods

Kula is one of the major coastal communities in Akuku-Toru Local Government Area of Rivers State, Nigeria. It is a swampy mangrove area located in the Niger Delta with geographical coordinates of latitude 4.35 and longitude 6.60, a few feet above sea level. Its estuaries, tributaries and creeks flows through Santa Barbara (Owuanga toru) San Batholomew (Aguda Toru) down to the Atlantic Ocean. Kula territory comprises many other settlements in different locations that are hosting the oil and gas flow stations mounted by Shell SPDC and Chevron.

Oil exploration and exploitation activities have been in Kula kingdom since 1958, with the production of two hundred thousand barrels of crude oil per day and five million cubic feet of gas daily.



Figure 1. Map Showing the Study Area



2.1 Sample Collection and Identification

Samples of the sea foods were randomly collected from communities in Kula Kingdom. Samples were wrapped in aluminum foil, packed in labeled polythene bags and transported to the laboratory for analysis. Samples were collected in the month of November 2017-May 2018.

Table 1. Identification Code and Classification of the Samples

S/N	Sea food	Identification code	English name	(kalabari/Ijaw)
1	Tympanotonos fuscatus	TF	Mud creeper periwinkle	isam
2	Crassostrea gigas	CG	Pacific oyster	<u>mgbe</u>
3	Busycon carica	BC	Whelk	ngolo

2.2 Determination of PAHs in the Samples

PAHs were determined in the sample according to established protocol by USEPA (1986). The PAHs determination was done using gas chromatography (GC) coupled with flame ionization detector (FID) (Hewlett Packard, Wilmington, DE, USA), powered with HP chemstation Rev. A09:01 (10206) software to identify and quantify compounds. The GC was programmed as follows: the inlet and injection temperature was set at $275^{\circ}C-310^{\circ}C$; fused silica column [30m*0.25µmfilmof HP-5(thickness)]; split injection was adopted with a split ratio of 8:1. The rubber septum used and volume injected was 1uL. The column temperature was programmed as follows: 65° C for 2min: $65 - 260^{\circ}$ C at 12° C /min: 260-320^oC at 15° C /min and maintained at 310° C for 8 min and oven temperature was set at 65° C. Nitrogen was used as carrier gas. The hydrogen and compressed air pressure was 30psi. A standard mixture of the 16 PAHs was obtained and subsequently used for the PAHs analysis. Comparison between the retention time of standards and that obtained from the extract of 1 ml was used to identified the compounds while individual PAHs analysis were used for the quantification. To ensure accuracy for all the PAHs measured, an analytical blank and spike sample consisting of all reagents were run with 6 samples to determine cross-contamination and interference. The efficiency of the analytical method used for the PAHs were assessed by recovery of internal standard.

Variables	Unit	values	Reference
Reference dose (RfD)	mg/kg/day	Table 2	USEPA (2008)
Fish ingestion rate (IR)	mg/kg/day	6500	Ihedioha et al. (2016)
Exposure duration (ED)	years	30	Tongo et al. (2015)
Adult body weight (BW)	kg	70	Orish et al. (2015)
Average life time for cancer effects (AT)	days	25500	Tongo et al. (2015)
Exposure frequency(EF)	day/year	365 (ingestion)	Tongo et al. (2015)
Cancer slope factor (CSF)	mg/kg.day	Table 2	USEPA (2008)
Carcinogenic potency of	mg/kg/day	7.3	Tongo et al. (2015)
benzo[a]pyrene(TEQ)(∑ BaPteq			
Toxicity equivalence factor (TEFi)	No unit	Table 7	Nisbet and LaGoy (1992)
Maximum acceptable risk (RL)	No unit	10 ⁻⁵	USEPA,2000

Table 2. Human Model Toxicological Variables for Assessment



PAHs	Code	Cancer slope	PAHs	Code	RfD
		Factor mg/kg.day			mg/kg/day
Benzo[a]anthracene	BaA	7.3×10^{-1}	Naphthalene	Nap	0.02
Chrysene	Chr	7.3×10^{-3}	2- Methylnaphthalene	2MNap	0.04
Benzo[k]fluoranthene	BkF	7.3×10^{-2}	Acenephthylene	Acy	0.02
Benzo[a]pyrene	BaP	7.3	Acenephthene	Ace	0.06
Benzo[b]floranthrene	BbF	7.3×10^{-1}	Fluorine	F lu	0.04
Indeno[1,2,3]pyrene	IDP	7.3×10^{-1}	Phenanthene	Phe	NA
Dibenzo[a,h]anthracene	DBA	7.3	Anthracene	Ant	0.3
			Fluoranthene	Fl n	0.04
			Pyrene	Pyr	0.03

Table 3. Cancer Slope Factor (CSf) and Reference Dose (RfD)

2.3 Exposure Assessment

2.3.1 The Estimated Daily Intake (EDI)

The Estimated Daily Intake (EDI) of PAHs via consumption of sea foods was assessed for adult population using Equation (1).

$$EDI = (C \times FIR) / (BW)$$
(1)

Where: EDI = Estimated daily intake (mg/kg/day)

The consumption rate for shell fish (sea food) was given as 6500mg/kg/day (Ihedioha *et al., 2016*).

2.4 Toxicity Assessment

2.4.1 Non Cancer Effects

Risks from exposure to PAHs through sea fish consumption was calculated as the hazard quotient (HQ) and hazard index (HI) using equations (2) and (3).

$$HQ = EDI /RfDi$$
 (2)

The total non-carcinogenic risk also known as hazard index was evaluated by the sum of HQ of each PAH using equation (3).

Hazard index (HI) =
$$HQ_1 + HQ_2 + HQ_3 + \dots + HQ_n$$
 (3)

Where: HQ = non-cancer hazard quotient.

RfD = chronic oral reference dose, which is an estimate of a daily oral exposure level for the human population.

2.4.2 Toxic Potency Assessment

Concentration of each PAH at the sample location was converted into Benzo[a] pyrene (Orish *et al.*, 2015). Toxicity equivalent (TEQ) method was used and calculated using equations (4).

B [a] Pteq. =
$$Cp \times TEFip$$
 (4)



Toxic Potency Assessment of PAHs in the sample environment was obtained by summing the toxic potencies of individual PAHs (B [a] Pteq) using equation (5).

$$(TEQs) = \Sigma B [a] Pteq$$
(5)

The toxic equivalent factor (TEFs) of the sixteen (16) PAHs values are presented in Table 7.

2.4.3 The Screening Value (SV)

SV is the threshold concentration of chemicals in edible tissue that is of potential public health concern (Tongo *et al.*, 2017).

The screening value was calculated using Equation (6)

$$SV = (RL/SFX BW)/IFR$$
 (6)

Where RL = Maximum acceptable risk level (10⁻⁵)

2.4.4 Four PAHs Index

It is the sum of four different poly-cyclic aromatic hydrocarbons, namely benzo[a]anthracene (B[a]A),chrysene (Chr), benzo[b]fluoranthene (B[b]FL), and benzo[a]pyrene(B[a]P) (Nwaichi and Ntorgbo, 2016., Tongo *et al.*, 2017).

4PAHs index was estimated using Equation (7).

$$PAH4 Index (PAH4) = (B[a]A + Chr + B[b]FL + B[a]P)$$
(7)

The maximum permissible level recommended is 30ug/kg (EU, 2014).

2.5 Excess Cancer Risk (ECR)

The excess cancer risk induced by dietary exposure to PAHs through sea food consumption was calculated using equation (8)

$$ECR = Q X B [a] Pteq X IFR X ED/(BW X ATn)$$
 (8)

2.6 Statistical Analysis

The data were statistically analyzed by SPSS software version 26. One-way ANOVA were applied for evaluating the significant difference between.

3. Results and Discussion

The mean concentration of each PAH is presented in Table4. The values ranged from 0.001-2.00 µg/kg, 0.002-0.689 µ/kg and 0-0.930 µ/kg in for whelk, oyster and periwinkle respectively. BaA concentrations were highest in whelk (0.689 \pm 0.003 µg/kg) and Periwinkle (0.930 \pm 0.001 µg/kg) while Napthalene concentration was highest in oyster (2.000 \pm 0.000 µ/kg). The total PAHs value obtained in (µ/kg) was highest for oyster (3.977 \pm 0.024), followed by whelk (1.797 \pm 0.013) and then periwinkle (1.564 \pm 0.017). The high values obtained may be attributed to poor metabolic clearance of PAHs in these marine organisms



and may be passed to human through the food chain with high degree of toxicity (Zelinkova and Wendi, 2015). PAHs that enter the marine environment via various means such illegal oil refininig popularly called bunkering or kpoo fire, atmospheric fallout, effluents from industrial and treatment plants and sewage, settle at sediments of estuaries and overtime bio-accumulate in these organisms that feed on sediments and filter large amount of water (Diacono and Montemurro, 2010).

S/N	PAHs	Code	Whelk	Oyster	Periwinkle
1	Naphthalene	Nap	0.002 ± 0.000	2.000±0.000	0.009 ± 0.000
2	2- Methylnaphthalene	2MNap	0.006 ± 0.000	0.345 ± 0.002	0.004 ± 0.000
3	Acenephthylene	Acy	0.048 ± 0.001	0.207 ± 0.001	-
4	Acenephthene	Ace	0.030 ± 0.001	0.073 ± 0.002	0.026±0.002
5	Fluorine	Flu	0.033 ± 0.003	0.025 ± 0.002	-
6	Phenanthene	Phe	0.160 ± 0.001	0.017 ± 0.003	0.014 ± 0.001
7	Anthracene	Ant	0.036 ± 0.004	0.026 ± 0.001	0.002±0.000
8	Fluoranthene	Fln	0.102 ± 0.002	0.252 ± 0.001	0.486±0.003
9	Pyrene	Pyr	0.273 ± 0.002	0.491 ± 0.003	-
10	Benzo[a]anthracene	BaA	0.689 ± 0.003	0.434 ± 0.001	0.930±0.001
11	Chrysene	Chr	0.079 ± 0.001	0.059 ± 0.001	-
12	Benzo[b]fluoranthene	BbF	0.280 ± 0.003	0.017 ± 0.002	-
13	Benzo[k]fluoranthene	BkF	0.04 ± 0.002	0.008 ± 0.000	0.014±0.002
14	Benzo[a] Pyrene	BaP	0.014 ± 0.004	$0.010\pm\!0.001$	0.016±0.003
15	Indeno[1,2,3]pyrene	IDP	0.003 ± 0.000	0.012 ± 0.002	0.005 ± 0.000
16	Dibenzo[a,h]anthracene	DBA	0.002 ± 0.000	0.001 ± 0.000	0.058 ± 0.001
	Total PAHs	Σ PAHs	1.797±0.013	3.977±0.024	1.564±0.017
	Total Carcinogenic PAHs	Σ CPAHs	0.418 ± 0.023	0.048 ± 0.010	0.093±0.014

Table 4. Mean Concentration of Each PAH in Selected shell fish (µg/kg)

The EDI in (mg/day) estimated from individual PAH via consumption of shell fish (whelk, oyster and periwinkle) is shown in Table 5. The values obtained ranged from 2.00×10^{-4} to 6.40×10^{-2} , 7.00×10^{-4} to 1.86×10^{-1} and 0 to 8.64×10^{-2} respectively which is far above the oral reference dose (RfD). The values obtained showed that daily consumption of seafood from these study sites could adversely affect human health ranging from neuronal and hepatocellular toxicity, peripheral gastrointestinal bleeding, vascular disease, oxidative stress.

Table 5. Estimated Daily Intake of PAHs in Selected Shell fish	(mg/kg/day)
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S/N	PAHs	Code	Whelk	Oyster	Periwinkle
1	Naphthalene	Nap	0.0002	0.1857	0.0008
2	2- Methylnaphthalene	2MNap	0.0005	0.0320	0.0004
3	Acenephthylene	Acy	0.0045	0.0192	-
4	Acenephthene	Ace	0.0028	0.0068	0.0024
5	Fluorine	Flu	0.0031	0.0023	-
6	Phenanthene	Phe	0.0149	0.0016	0.0013
7	Anthracene	Ant	0.0031	0.0024	0.0002
8	Fluoranthene	Fln	0.0095	0.0234	0.045
9	Pyrene	Pyr	0.0254	0.0456	
10	Benzo[a]anthracene	BaA	0.0640	0.0403	0.0864
11	Chrysene	Chr	0.0073	0.0055	-
12	Benzo[b]fluoranthene	BbF	0.0260	0.0016	-



13	Benzo[k]fluoranthene	BkF	0.0038	0.0007	0.0013
14	Benzo[a] Pyrene	BaP	0.0014	0.0010	0.0015
15	Indeno[1,2,3]pyrene	IDP	0.0003	0.0011	0.0005
16	Dibenzo[a,h]anthracene	DBA	0.0002	0.0012	0.0054
	Sum of EDI	Σ EDI	0.1669	0.3704	0.1451

Potential risk to human health through more than one pollutant was measured by the hazard index (HI) which is given in Table 6. HI is the sum of all HQs calculated for individual PAH. A value of HQ or HI < 1 implies no significant non-cancer risks (no hazard); a value ≥ 1 implies significant non-cancer risks (hazard), which increases with increasing value of HQ or HI (Wei, *et al.*, 2015). The values observed for the HI, via ingestion from all the sample is greater than 1 which shows that levels of PAHs in the sample have potential non-carcinogenic adverse health risk.

S/N	PAHs	Code	Whelk	Oyster	Periwinkle
1	Naphthalene	Nap	0.0010	9.2850	0.0400
2	2- Methylnaphthalene	2MNap	0.0125	0.8000	0.0100
3	Acenephthylene	Acy	0.2250	0.9600	NA
4	Acenephthene	Ace	0.0467	0.1133	0.0400
5	Fluorine	Flu	0.0775	0.0575	NA
6	Phenanthene	Phe	NA	NA	NA
7	Anthracene	Ant	0.0103	0.0080	0.0007
8	Fluoranthene	Fln	0.2375	0.5850	1.1250
9	Pyrene	Pyr	0.8467	1.5200	NA
10	Benzo[a]anthracene	BaA	NA	NA	NA
11	Chrysene	Chr	NA	NA	NA
12	Benzo[b]fluoranthene	BbF	NA	NA	NA
13	Benzo[k]fluoranthene	BkF	NA	NA	NA
14	Benzo[a] Pyrene	BaP	NA	NA	NA
15	Indeno[1,2,3]pyrene	IDP	NA	NA	NA
16	Dibenzo[a,h]anthracene	DBA	NA	NA	NA
	Hazard Index	ΣHQs	1.4572	13.3288	1.2157

Table 6. HQ values (Non-carcinogenic Effects) of PAHs in Selected Shell fish

In recent studies, specific criteria have been established for a safe level of pollutants in shellfish for human consumption (Tongo *et al.*, 2017). The toxic equivalents (TEQs) values (mg/kg) were 1.276×10^{-4} , 1.252×10^{-4} and 4.034×10^{-4} for whelk, oyster and periwinkle respectively these values were significantly (p<0.05) higher than the screening value (SV) for shellfish 1.81×10^{-5} mg/kg. This is indication of increased carcinogenic potential of total PAHs via consumption of these sea foods or shell fish as shown in Tables 7 and 8.

Table 7. Carcinogenic potencies (B(A)Pteq) of PAHs in the Selected Shell fish

S/N	PAHs	Code	Whelk	Oyster	Periwinkle	TEF
1	Naphthalene	Nap	2.24E-09	2.00E-06	8.80E-09	0.001
2	2- Methylnaphthalene	2MNap	5.84E-09	3.50E-07	4.00E-09	0.001
3	Acenephthylene	Acy	4.82E-08	2.10E-07	-	0.001
4	Acenephthene	Ace	3.02E-08	7.30E-08	2.60E-08	0.001
5	Fluorine	Flu	3.33E-08	2.50E-08	-	0.001
6	Phenanthene	Phe	1.60E-07	1.70E-08	1.40E-08	0.001



7	Anthracene	Ant	3.36E-07	2.60E-07	2.40E-08	0.01
8	Fluoranthene	Fln	1.02E-07	2.50E-07	4.90E-07	0.001
9	Pyrene	Pyr	2.73E-07	4.90E-07	-	0.001
10	Benzo[a]anthracene	BaA	6.89E-05	4.30E-05	9.30E-05	0.1
11	Chrysene	Chr	7.90E-07	6.00E-07	-	0.01
12	Benzo[b]fluoranthene	BbF	2.80E-05	1.70E-06	-	0.1
13	Benzo[k]fluoranthene	BkF	4.07E-06	7.80E-07	1.40E-06	0.01
14	Benzo[a] Pyrene	BaP	1.48E-05	1.00E-05	1.60E-05	1
15	Indeno[1,2,3]pyrene	IDP	3.39E-07	1.20E-06	5.30E-07	0.1
16	Dibenzo[a,h]anthracene	DBA	9.75E-06	6.40E-05	2.90E-04	5
	$\Sigma B(A)$ Pteq	TEQs	1.28E-04	1.25E-04	4.03E-04	

Table 8. Screening value for Six (6) Carcinogenic PAHs in the Selected Shellfish

S/N	PAHs	Code	Screeningvalue (SV)
1	Benzo[a]anthracene	BaA	1.5×10^{-7}
2	Chrysene	Chr	1.4×10^{-5}
3	Benzo[b]fluoranthene	BbF	1.48x10 ⁻⁶
4	Benzo[k]fluoranthene	BkF	1.48x10 ⁻⁶
5	Benzo[a] Pyrene	BaP	1.0×10^{-8}
6	Indeno[1,2,3]pyrene	IDP	1.5x10 ⁻⁷
	Total SV	Σ SV	1.81x10 ⁻⁵

The sum of four different poly-cyclic aromatic hydrocarbons, namely benzo[a]anthracene (B[a]A),chrysene (Chr), benzo[b]fluoranthene (B[b]FL), and benzo[a]pyrene(B[a]P) has also been used to establish the occurrence and toxicity of PAHs in food (EFSA, 2008, Nwaichi and Ntorgbo, 2016). The values of Σ 4PAHs obtained in the sea foods were $1x10^{-3}$, $5.2x10^{-4}$ and $9.46x10^{-4}$ mg/kg for whelk, oyster and periwinkle respectively. These observed values were below the maximum permissible level recommended by EU (30μ /kg). This implies that the four PAHs could not pose potential carcinogen health effects to humans via consumption as presented in Table 9.

	C/NI	DATE.	C. 1.	3371 11.	0	D
Table 9. R	isk of Fou	r PAHs (ΣPAHs) in t	he Selecte	ed Shellf	ĩsh (μg/l	kg)

S/N	PAHs	Code	Whelk	Oyster	Periwinkle
1	Benzo[a]anthracene	BaA	0.689	0.434	0.93
2	Chrysene	Chr	0.079	0.059	-
3	Benzo[b]fluoranthene	BbF	0.28	0.017	-
4	Benzo[a] Pyrene	BaP	0.014	0.01	0.016
	Total PAHs	Σ4PAHs	1.062	0.52	0.946

The result for the excess cancer risk (ECR) is presented in Table 10. All carcinogenic PAHs which include Chrysene, Benzo [a] anthracene, Benzo [k] fluoranthene, Benzo [a] pyrene, Indeno [1, 2, 3-c, d] pyrene, Benzo [b] fluoranthene and Dibenzo [a, h] anthracene were detected in whelk and oyster except periwinkle. Total Carcinogenic PAHs (μ g/kg) obtained for whelk oyster and periwinkle were (0.418±0.023), (0.048±0.10), and (0.093±0.14) respectively. The estimated excess cancer risk (ECR) obtained for whelk was (3.0x10⁻⁴), oyster (2.00x10⁻⁴) and periwinkle (3.24x10⁻⁴). These values were far above the USEPA acceptable (1x10⁻⁴). Thus pose carcinogenic risk that may be due to PAHs burden from the environment.



S/N	PAHs	Code	Whelk	Oyster	Periwinkle
1	Naphthalene	Nap	5.20E-09	4.64E-06	2.04E-08
2	2- Methylnaphthalene	2MNap	1.35E-08	8.12E-08	9.28E-09
3	Acenephthylene	Acy	1.12E-07	4.87E-07	
4	Acenephthene	Ace	7.01E-08	1.70E-07	6.00E-08
5	Fluorine	Flu	7.73E-08	5.80E-08	
6	Phenanthene	Phe	3.71E-07	4.00E-08	3.22E-08
7	Anthracene	Ant	7.80E-07	6.00E-08	5.67E-08
8	Fluoranthene	Fln	2.37E-08	5.80E-08	1.14E-07
9	Pyrene	Pyr	6.33E-07	1.37E-07	-
10	Benzo[a]anthracene	BaA	1.60E-04	1.00E-06	2.16E-04
11	Chrysene	Chr	1.83E-06	1.40E-07	-
12	Benzo[b]fluoranthene	BbF	6.50E-05	4.00E-07	-
13	Benzo[k]fluoranthene	BkF	9.44E-06	1.81E-08	3.25E-06
14	Benzo[a] Pyrene	BaP	3.43E-05	2.32E-07	3.71E-05
15	Indeno[1,2,3]pyrene	IDP	7.86E-07	2.78E-07	1.23E-07
16	Dibenzo[a,h]anthracene	DBA	2.26E-05	1.50E-04	6.73E-07
	Excess cancer risk	ΣECR	3.00E-04	2.00E-04	3.24E-04

Table 10. Excess cancer risk (ECR) of PAHs in the Selected Shellfish

4. Conclusion

PAHs are major pollutant of the aquatic environment that may settle at sediments of coastal estuaries and over time may bio-accumulate in these sea organisms and finally passed on to humans through the food chain with high degree of toxicity. Results obtained from this study confirms the rapid anthropogenic activities going on in the study region such as illegal oil refining known as kpoo fire or bunkering and have contributed to the increased availability of PAHs. The daily consumption of shellfish constitutes a potential health hazard. Carcinogenic indices indicated PAHs contaminated shellfish exposure the local populace to cancer risks. In the light of the above findings, there is need for policymakers and other concerned stakeholders to regulate anthropogenic activities that may result to increase emission of PAHs in the study area and protect local residents from impeding health risk associated with exposure.

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