

Seasonal Dynamics of the Microbial and Physicochemical Characteristics of Streams and Boreholes in Uzuakoli, Eastern Nigeria

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 Received: February 10, 2019
 Accepted: March 3,, 2019

 doi:10.5296/jbls.v10i2.14456
 URL: https://doi.org/10.5296/jbls.v10i2.14456

Abstract

Water samples from twenty water sources (fifteen boreholes and five streams) in Uzuakoli, Nigeria were collected for the period of 6 months covering the dry and rainy seasons to assess the level of contamination. The Microbiological characteristics including heterotrophic counts, coliform counts and physicochemical parameters includes pH, turbidity, dissolved oxygen, calcium, potassium, nitrate, magnesium and phosphate were evaluated using standard methods. The total Heterotrophic counts for the borehole during the dry and rainy season were 8.3×10^3 cfu/ ml and 10.8 x 10^4 cfu/ ml. The Heterotrophic counts for the stream were 12.7 x 10^4 cfu/ ml and 17.8x 10⁶ cfu/ ml. The frequency of occurrence of the isolates are *Staphylococcus aureus* 63% in borehole and 85% in streams, Pseudomonas aeruginosa 49% in boreholes and 95% in streams, Proteus sp 52% in boreholes and 97% in streams, Streptococcus sp 46% in boreholes and 53% in streams, Enterobacter aerogenes 33% in boreholes and 63% in streams, Escherichia coli 16% in boreholes and 53% in streams and Salmonella sp no percentage in boreholes and 40% in streams. The result shows a significant difference at (P≤0.05) for the bacterial isolates. The physicochemical parameters of the borehole and stream water samples during the dry and rainy seasons were determined. The temperature ranged from 25°C 32°C; pH ranged from 5.3 8.1; turbidity ranged 0.03 3.23; dissolved oxygen ranged from 3.45–7.40mg/l; biochemical oxygen demand ranged from 1.20–4.32mg/l; chemical oxygen demand ranged from 2.50-5.21mg/l; Calcium ranged from 0.81-5.64mg/l; potassium ranged from 1.01-4.22mg/l; Nitrate ranged from 1.49-4.02mg/l; magnesium ranged from 0.13–2.20mg/l; phosphate ranged from 0.51–2.01mg/l. The water samples were all within the WHO limits apart from sample from Iyi Agbozu that had temperature of 32°C.



Keywords: microbial, physicochemical characteristics, seasonal dynamics, eastern Nigeria

1. Introduction

Water is indispensably and intricately connected to life without which there is no life. This is the reason for which water must be given the necessary attention at all times. The supply of safe drinking water to all has therefore engaged the attention of many individual, groups, governmental organization and private (Wahab et al., 2012). Natural water is never absolutely pure, as it carries traces of other substances which bestow on it physical, chemical and bacteriological characteristics. The nature and amount of these substances called impurities vary with sources of the water (Oyhakilome et al., 2012). In developing countries including Nigeria, the majority of people live in rural areas. Water from rivers, streams, wells and more recently boreholes serve as the main sources of water for drinking and domestic use (Ibe and Okplenye, 2005). Conformation with physiochemical and microbiological standards is of special interest because of the capacity of water to spread diseases within a large population. A good knowledge of the chemical qualities of raw water is necessary so as to guide its suitability for use (Sunday and Innocent 2012). The presence of these biological and chemical parameters in drinking water affects the pH, colour, salinity, dissolved oxygen, total solutes, alkalinity, hardness and conductivity of the water beyond WHO specified tolerable limits. This leads to various water borne diseases including diarrhea, cholera, typhoid fever, shigellosis, giardiasis, schistosomiasis, hepatitis and onchocerciasis (Akubuenyi et al., 2013).

2. Materials and Methods

Study Area

Uzuakoli is in Bende Local Government area, of Abia State and has a long history dating back to the time of the slave trade when its market, Agbo Agwu, was a major centre for slave exchange. It is located in the northern region of Abia State. Uzuakoli lies between Latitude 5.6333 and Longitude 7.5667. The community is made up of five villages, Agbozu, Amamba, Amankwo, Eluama and Ngwu, each of the villages have their streams and boreholes.

Sample Collection

The sample were collected twice a month for a period of six months, covering February, march, april for dry season and june, july and august for rainy season. Water samples were collected using sterile 500ml containers which were first washed and properly sterilized to avoid contamination. The stream water samples were collected by unscrewing the cap of the container and holding the container near its base in the hand and plunging its neck downwards below the surface. The containers were turned until neck points slightly upwards and mouth is directed towards the current. When the water fills the containers it was carefully removed and corked. In other to collect sample from borehole, cotton wool soaked in ethanol was used to disinfect the nozzle of the boreholes and then the tap was turned on to allow water to run for two minutes before sterile 500ml screw capped plastic containers were carefully uncapped and filled with water and recapped. The samples were labeled with code names for proper identification. Thereafter the water samples were transported to the laboratory for analysis within six hours of collection (Cheesbrough, 2010).



Enumeration of Total Heterotrophic Bacteria Count

Samples of the stream water samples were serially diluted in ten folds. Total heterotrophic counts were determined using pour plate technique. The molten nutrient agar, were poured into the Petri dishes containing 1.0mL of the dilution for the isolation of the total heterotrophic bacteria. They plates were swirled to mix thoroughly and the colony counts were taken after incubating the plates at room temperature for 48h and preserved by sub-culturing the bacterial isolates into nutrient agar slants which were used for biochemical tests (APHA, 1998).

Characterization and Identification of Bacterial

Bacterial isolates were characterized and identified after studying the Gram reaction as well as cell morphology. Other tests performed were spore formation, motility, oxidase and catalase production, citrate utilization, oxidative/fermentation (O/F) utilization of glucose; indole and coagulase production, starch hydrolysis, sugar fermentation, methyl red-Voges Proskaur reaction and urease production. The tests were performed according to the methods of (Cheesbrough, 2010). Microbial identification was performed using the keys provided in the *Bergeys Manual of Determinative Bacteriology* (1994).

Physicochemical Analysis

The physicochemical parameters include temperature, pH, dissolved oxygen (DO), turbidity, nitrate, phosphate, calcium, potassium, biochemical oxygen demand (BOD) and chemical oxygen demand (COD). The pH was measured in-situ using pH meter; the temperature was measured in situ using mercury in bulb thermometer in centigrade scale, turbidity was determined using spectrophotometric method. Potassium, magnesium, phosphate, nitrate, dissolved oxygen, Chemical oxygen demand and biochemical oxygen demand were determined by method of (ALPHA 1998).

3. Results

Figures 4.1 showed the seasonal variation of the total heterotrophic bacterial count. Table 4.1 showed the frequency of occurrence of bacteria. The physicochemical parameters are shown in table 4.2 - 4.3.





Figure 4.1. Mean Seasonal Heterotrophic Counts in Borehole and Stream Water Samples

(cfu/ml)

Bacterial Isolates	Isolates in borehole samples (n=180)	Isolates in stream samples (n=60)					
Staphylococcus aureus	113 (63%)	51 (85%)					
Streptococcus sp	83 (46%)	32 (53%)					
Escherichia coli	29 (16%)	32 (53%)					
Salmonella sp	0 (0%)	24 (40%)					
Pseudomonas aeruginosa	89 (49%)	57 (95%)					
Enterebacter aerogenes	59 (33%)	38 (63%)					
Proteus sp	94 (52%)	58 (97%)					



Table 4.2. Physicochemical parameters of borehole water samples during dry and rainy season

	Mean of Dry season							Mean				
Parameters	Agbozu	Amamba	Amankwo	Eluoma	Ngwu		Agbozu	Amamba	Amankwo	Eluoma	Ngwu	WHO Standards (2004)
pН	5.5	6.1	6.7	5.5	5.9		6.2	5.8	6.3	6.6	7.8	6.5 - 8.5
Temp (°C)	27	28	29	28	30		25	25	26	28	29	25°C - 30°C
Turbidity	0.04	0.04	0.03	0.06	0.06		0.36	2.21	1.68	2.21	1.10	5
DO (mg/l)	4.03	4.92	3.82	5.01	3.45		6.43	6.01	7.31	7.31	7.40	14
BOD(mg/l)	2.35	2.39	2.31	1.20	2.42		2.40	2.89	3.61	3.81	3.90	< 4
COD(mg/l)	3.26	3.05	3.26	2.50	3.01		3.22	3.50	3.01	2.92	3.33	< 10
Ca ²⁺ (mg/l)	0.81	2.53	3.50	1.25	2.53		2.41	2.22	2.41	0.82	3.19	50
K ⁺ (mg/l)	3.43	3.41	2.45	3.43	2.56		4.22	2.71	3.25	1.01	3.25	-
P (mg/l)	0.51	1.22	1.49	0.62	2.01		0.23	0.11	1.01	1.88	1.43	-
Mg ²⁺ (mg/l)	0.22	0.71	1.30	1.23	1.23		0.14	0.62	1.13	1.14	1.14	30
$NO_3 (mg/l)$	3.96	3.66	3.02	3.28	3.34		2.45	4.02	3.20	3.21	4.02	10

Table 4.3. Physicochemical parameters of stream water samples during dry and rainy season

Mean of Dry season							Mean of Rainy season						
Parameters	Iyi Agbozu	Ogbitiamapu	Iyi Amankwo	Iyi Nzu	Geregere Ngwu		Iyi Agbozu	Ogbitiamapu	Iyi Amankwo	Iyi Nzu	Geregere Ngwu	WHO Standards (2004)	
pH	7.6	6.8	7.4	6.9	7.6		7.4	7.6	7.9	7.6	8.1	6.5 - 8.5	
Temp (°C)	32	28	30	29	29		28	26	28	27	28	25°C - 30°C	
Turbidity	0.14	0.08	0.14	0.12	0.11		3.23	1.60	3.23	1.87	1.60	5	
DO (mg/l)	5.91	6.01	5.91	6.01	6.45		5.40	6.01	5.41	6.43	6.01	14	
BOD (mg/l)	3.80	3.62	3.30	2.30	3.12		4.32	3.81	2.89	4.32	3.61	< 4	
COD (mg/l)	4.20	5.21	5.21	4.23	5.01		4.31	3.94	3.56	4.11	3.81	< 10	
$Ca^{2+}(mg/l)$	5.64	4.32	4.32	2.48	3.33		2.31	2.42	3.29	3.29	2.64	50	
K ⁺ (mg/l)	1.08	2.02	1.52	1.08	2.31		3.52	3.02	1.29	2.31	3.91	-	
P (mg/l)	1.78	1.78	2.32	2.01	1.92		0.89	1.24	1.80	2.01	1.42	-	
Mg ²⁺ (mg/l)	1.62	2.30	1.62	1.43	2.01		0.91	2.20	1.73	1.66	1.31	30	
$NO_3 (mg/l)$	1.49	2.13	1.52	1.49	2.02		2.13	1.52	2.11	2.44	1.82	10	

5. Discussion

The microbiological analysis reveals that the microbial load varied from dry season rainy season across the streams and boreholes. The borehole waters had the least microbial counts when compared to the stream waters. The highest microbial counts were recorded in streams during the rainy season which could be due to runoffs, domestic and human activities. However the bacterial load decreased downstream and this could be attributed to possible self-purification and dilution of the pollutants (Eze *et al.*, 2013). The isolated bacteria were *Staphylococcus aureus, Pseudomonas aeruginosa, Proteus sp, Streptococcus sp, Enterobacter aerogenes, Escherichia coli* and *Salmonella sp.* The borehole samples are free of *Salmonella*



sp. unlike the streams samples. These organisms are important human pathogens associated with a variety of infectious diseases such as gastroenteritis, typhoid fever, dysentery, cholera and urinary tract infections etc. Their presence raises serious public health concern because they are known causative agents of many water borne diseases and indicates that these water sources are not potable (Akubuenyi *et al.*, 2013). The presence of *Escherichia coli* which is the most common indicator of faecal pollution in a water sample is an indication of the presence of enteric pathogens. This confirms the result of an earlier study by Okezie and Nwachukwu (2018), indicating that most boreholes and streams are heavily contaminated with faecal matter.

The physicochemical parameters analyzed were all within the WHO limit for drinking water except for the temperature and BOD for the streams. The stream water samples had pH within the WHO permissible limits of 6.5 - 8.5 while borehole water samples from Agbozu, Eluoma and Ngwu had pH values below the WHO permissible limits. The low pH values obtained from the borehole samples could be attributed to dissolved mineral salts in water (Shittu *et al.*, 2008). The temperature of the samples differed significantly. The highest was observed in the stream samples from $26^{\circ}C - 32^{\circ}C$ and lowest in the borehole samples from $25^{\circ}C - 30^{\circ}C$. However, temperature of a water body is affected by a number of factors such as climate of the area, extent of shade from direct sunlight and depth of water. The drop in temperature across the months in this study could be as a result of increased rainfall, inflow of runoffs into water bodies and cooled weather (Ekhaise and Anyansi, 2005). All the samples had turbidity values within the WHO permissible limits. Turbidity could be caused due to runoff and human activities around the streams. Excessive turbidity in water causes problems with water purification process (John *et al.*, 2008).

Dissolved oxygen is one of the important and critical characteristics of water quality assessment. During the dry season the stream had more dissolve oxygen values than the boreholes while during the rainy season the boreholes had more values than the stream. The low values observed may be as a result of the increased runoffs of agricultural wastes and industrial effluents discharged into the drains that place high demand on the dissolve oxygen (Shittu *et al.*, 2008).

Biochemical oxygen demand (BOD) measures the amount of dissolved oxygen needed by microorganisms to break down organic matter present in a water samples over a specific time (Anake *et al.*, 2013). According to Oluyemi *et al.* (2010) BOD values less than 4 mg/l suggests that the water is less polluted by organic matter and could support aquatic life. The BOD for the borehole and stream samples is below 4 mg/l apart from BOD from Iyi Agbozu and Iyi Nzu. The stream water samples had more BOD values than the borehole samples. However, comparing the BOD value for dry season and rainy season, the rainy season had the highest BOD values.

Chemical Oxygen Demand value for the borehole samples is low compared to the stream samples. The borehole and stream water samples have COD values below 10 mg/l.

Calcium ions measured in the borehole and stream water samples were within the WHO permissible limit. Excess calcium in water could causes hardness of water. Calcium is



essential and helps in bone formation. It is commonly present in all water bodies where it usually comes from the leaching of rocks (Akubuenyi et al., 2013).

Phosphate, Potassium and Nitrate were all within the WHO permissible limits for drinking water. Phosphates and nitrates are important ingredients in plant blooms and eutrophication of streams (WHO, 2006). Nitrate in concentration greater than 45mg/l is undesirable in domestic water supplies because of the potential toxic effect on young infants. Methemoglobinemia is a disease caused by nitrate, which occurs when it is converted to nitrite in the intestines. Nitrate cannot be removed from water by boiling but must be treated by distillation (Sunday and Innocent, 2012).

6. Conclusion

There is seasonal variation in the microbial load of the water samples especially, during the rainy season which could be as a result of runoffs and human activities. This may lead to spread of diseases.

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